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Reflex FORS2 Spectropolarimetry Tutorial

VLT-MAN-ESO-19500-....

Issue 1.0

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1 Introduction And Scope

Reflex is the ESO Recipe Flexible Execution Workbench, an environment to run ESO VLT pipelines which employs a workflow engine to provide a real-time visual representation of a data reduction cascade, called a workflow, which can be easily understood by most astronomers. The basic philosophy and concepts of Reflex have been discussed by Freudling et al. (2013A&A...559A..96F). Please reference this article if you use Reflex in a scientific publication.

Reflex and the data reduction workflows have been developed by ESO and instrument consortia and they are fully supported. If you have any issue, please contact usd-help@eso.org for further support.

This document is a tutorial designed to enable the user to employ the FORS2 workflow to reduce his/her data in a user-friendly way, concentrating on high-level issues such as data reduction quality and signal-to-noise (S/N) optimisation.

A workflow accepts science and calibration data, as delivered to PIs in the form of PI-Packs (until October 2011) or downloaded from the archive using the CalSelector tool¹ and organises them into DataSets, where each DataSet contains one science object observation (possibly consisting of several science files) and all associated raw and static calibrations required for a successful data reduction. The data organisation process is fully automatic, which is a major time-saving feature provided by the software. The DataSets selected by the user for reduction are fed to the workflow which executes the relevant pipeline recipes (or stages) in the correct order. Full control of the various recipe parameters is available within the workflow, and the workflow deals automatically with optional recipe inputs via built-in conditional branches. Additionally, the workflow stores the reduced final data products in a logically organised directory structure employing user-configurable file names.

The FORS2 Reflex workflow described in this tutorial supports the reduction of FORS2 spectropolarimetric observations (PMOS). The user is referred to the FORS2 user manual (Saviane, 2012² for more information on the instrument itself, and the FORS2 pipeline user manual (Izzo et al. 2012³) for the details of the spectroscopic FORS2 pipeline recipes. There are also other tutorials that guide you through the MOS/MXU/LSS workflow and the PMOS workflow respectively. Check the ESO pipeline main webpage.

The quick start section (see Section 4) describes the minimum effort to get started, and it makes up only two pages of text in this tutorial.

¹<http://www.eso.org/sci/archive/calselectorInfo.html>

²available at

<http://www.eso.org/sci/facilities/paranal/instruments/fors2/doc>

³available at <ftp://ftp.eso.org/pub/dfs/pipelines/fors/fors-pipeline-manual-5.2.pdf>

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2 Software Installation

The software pre-requisites for Reflex 2.8 may be found at:

http://www.eso.org/sci/software/pipelines/reflex_workflows

To install the Reflex 2.8 software and demo data, please follow these instructions:

1. From any directory, download the installation script:

```
wget ftp://ftp.eso.org/pub/dfs/reflex/install_esoreflex
```

2. Make the installation script executable:

```
chmod u+x install_esoreflex
```

3. Execute the installation script:

```
./install_esoreflex
```

and the script will ask you to specify three directories: the download directory <download_dir>, the software installation directory <install_dir>, and the directory to be used to store the demo data <data_dir>. If you do not specify these directories, then the installation script will create them in the current directory with default names.

4. You will be asked whether you want to use your Internet connection. Unless you want to reuse already downloaded packages (only advanced users), use the default Yes.
5. You will be given a choice of pipelines (with the corresponding workflows) to install. Please specify the numbers for the pipelines you require, separated by a space, or type “A” for all pipelines.
6. For the pipelines to be installed you will be prompted for the demo data sets to be installed. Type “A” for all demo datasets. Take into account that if you are installing in a directory that already contains data, it won’t be removed.
7. The script will also detect whether previous versions of the workflows or Reflex were installed and in this case you have the option to update links or remove obsolete cache directories. It is advised to use the defaults.
8. To start Reflex, issue the command:

```
<install_dir>/bin/esoreflex
```

It may also be desirable to set up an alias command for starting the Reflex software, using the shell command `alias`. Alternatively, the `PATH` variable can be updated to contain the <install_dir>/bin directory.

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3 Demo Data

Together with the pipeline you will also receive a demo data set, that allows you to run the `Reflex FORS2` workflow without any changes in parameters. This way you have a data set to experiment with before you start to work on your own data.

Note that you will need a minimum of ~ 0.5 GB, ~ 0.6 GB and ~ 1 GB of free disk space for the directories `<download_dir>`, `<install_dir>` and `<data_dir>`, respectively. The FORS2 demo data have been retrieved with the CalSelector tool⁴.

⁴<http://www.eso.org/sci/archive/calselectorInfo.html>

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4 Quick Start: Reducing The Demo Data

For the user who is keen on starting reductions without being distracted by detailed documentation, we describe the steps to be performed to reduce the science data provided in the FORS2 demo data set supplied with the Reflex 2.8 release. By following these steps, the user should have enough information to attempt a reduction of his/her own data without any further reading:



Figure 4.1: *The empty Reflex canvas.*

1. Start the Reflex application:


```
esoreflex &
```

The empty Reflex canvas as shown in Figure 4.1 will appear.

2. Now open the FORS2 workflow by clicking on **File** -> **Open File**, selecting first `fors-5.2.1` and then the file `fors_spec.xml` in the file browser. You will be presented with the workflow canvas shown in Figure 4.2. Note that the workflow will appear as a canvas in a new window.
3. To aid in the visual tracking of the reduction cascade, it is advisable to use component (or actor) highlighting. Click on **Tools** -> **Animate at Runtime**, enter the number of milliseconds representing the animation interval (100 ms is recommended), and click **OK**.
4. Under “Setup Directories” in the workflow canvas there are seven parameters that specify important directories (green dots). Setting the value of `ROOT_DATA_DIR` is the only necessary modification if you want to process data other than the demo data⁵, since the value of this parameter specifies the working directory within which the other directories are organised. Double-click on the parameter `ROOT_DATA_DIR` and a pop-up window will appear allowing you to modify the directory string, which you may either edit directly, or use the **Browse** button to select the directory from a file browser. When you have finished, click **OK** to save your changes.

⁵If you used the install script `install_esoreflex`, then the value of the parameter `ROOT_DATA_DIR` will already be set correctly to the directory where the demo data was downloaded.

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5. Click the  button to start the workflow
6. The workflow will highlight the `Data Organiser` actor which recursively scans the raw data directory (specified by the parameter `RAWDATA_DIR` under “Setup Directories” in the workflow canvas) and constructs the `DataSets`. Note that the raw and static calibration data must be present either in `RAWDATA_DIR` or in `CALIB_DATA_DIR`, otherwise `DataSets` may be incomplete and cannot be processed. However, if the same reference file was downloaded twice in different places this creates a problem as `Reflex` cannot decide which one to use.
7. The `Data Set Chooser` actor will be highlighted next and will display a “Select Datasets” window (see Figure 4.3) that lists the `DataSets` along with the values of a selection of useful header keywords⁶. The first column consists of a set of tick boxes which allow the user to select the `DataSets` to be processed. By default all complete `DataSets` which have not yet been reduced will be selected.
8. Click the `Continue` button and watch the progress of the workflow by following the red highlighting of the actors. A window will show which `DataSet` is currently being processed.
9. When the reduction of the current `DataSet` finishes, a pop-up window called *Product Explorer* will appear showing the datasets which have been so far reduced together with the list of final products. This actor allows the user to inspect the final data products, as well as to search and inspect the input data used to create any of the products of the workflow. Figure 4.4 shows the *Product Explorer* window.
10. The workflow will continue with the remaining `DataSets` following the same steps described above.
11. After the workflow has finished, all the products from all the `DataSets` can be found in a directory under `END_PRODUCTS_DIR` with the named with the workflow start timestamp. Further subdirectories will be found with the name of each `DataSet`.

Well done! You have successfully completed the quick start section and you should be able to use this knowledge to reduce your own data. However, there are many interesting features of `Reflex` and the FORS2 workflow that merit a look at the rest of this tutorial.

⁶The keywords listed can be changed by right-clicking on the `DataOrganiser Actor`, selecting `Configure Actor`, and then changing the list of keywords in the second line of the pop-up window.

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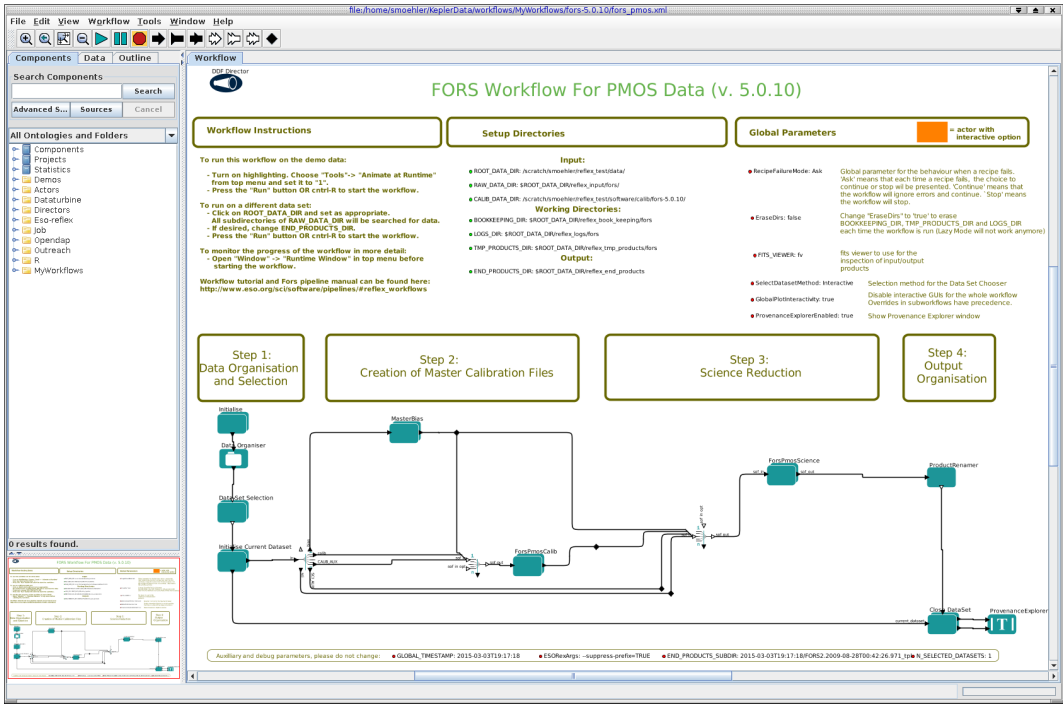


Figure 4.2: FORS2 workflow general layout.

The 'Select Datasets' pop-up window displays a table with the following data:

Selected	Data Set	#Files	OBS.TARG.NAME	CPL.TYPE	INS.OPT4.NAME
<input checked="" type="checkbox"/>	FORS2.2009-08-26T00:42:26.870_tpl	19	WD.2038-202	OBJECT	RET2
<input checked="" type="checkbox"/>	FORS2.2015-02-26T07:56:45.429_tpl	37	WD.1620-391	STD	RET2

Buttons at the bottom: Select complete, Select all, Deselect all, Save all, Inspect highlighted, Continue, Stop.

Figure 4.3: The “Select Datasets” pop-up window.

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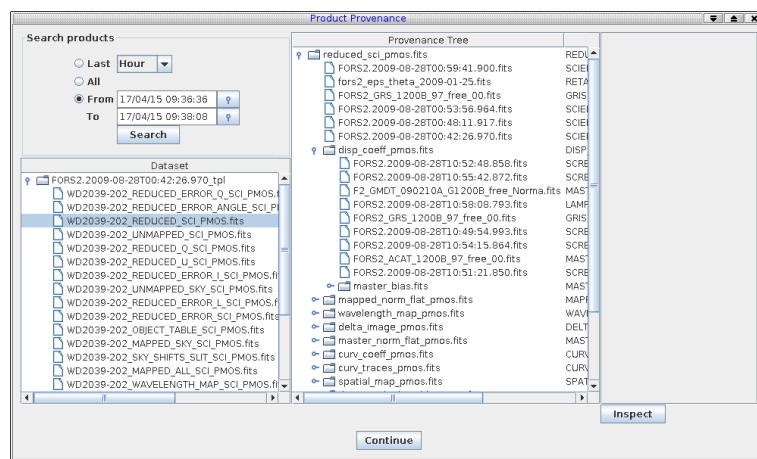


Figure 4.4: The Product Explorer shows all datasets reduced in previous executions together with the full reduction chain for all the pipeline products.

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5 About The Reflex Canvas

5.1 Saving And Loading Workflows

In the course of your data reductions, it is likely that you will customise the workflow for various data sets, even if this simply consists of editing the `ROOT_DATA_DIR` to a different value for each data set. Whenever you modify a workflow in any way, you have the option of saving the modified version to an XML file using `File -> Export As` (which will also open a new workflow canvas corresponding to the saved file). The saved workflow may be opened in subsequent Reflex sessions using `File -> Open`. Saving the workflow in the default Kepler format (.kar) is only advised if you do not plan to use the workflow in another computer.

5.2 Buttons

At the top of the `Reflex` canvas are a set of buttons which have the following useful functions:

-  - Zoom in.
-  - Reset the zoom to 100%.
-  - Zoom the workflow to fit the current window size (Recommended).
-  - Zoom out.
-  - Run (or resume) the workflow.
-  - Pause the workflow execution.
-  - Stop the workflow execution.

The remainder of the buttons (not shown here) are not relevant to the workflow execution.

5.3 Workflow States

A workflow may only be in one of three states: executing, paused, or stopped. These states are indicated by the yellow highlighting of the , , and  buttons, respectively. A workflow is executed by clicking the  button. Subsequently the workflow and any running pipeline recipe may be stopped immediately by clicking the  button, or the workflow may be paused by clicking the  button which will allow the current actor/recipe to finish execution before the workflow is actually paused. After pausing, the workflow may be resumed by clicking the  button again.

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6 The FORS2 Workflow

The FORS2 workflow canvas is organised into a number of areas. From top-left to top-right you will find general workflow instructions, directory parameters, and global parameters. In the middle row you will find five boxes describing the workflow general processing steps in order from left to right, and below this the workflow actors themselves are organised following the workflow general steps.

6.1 Workflow Canvas Parameters

The workflow canvas displays a number of parameters that may be set by the user. Under “Setup Directories” the user is only required to set the `RAWDATA_DIR` to the working directory for the `DataSet(s)` to be reduced, which, by default, is set to the directory containing the demo data. The `RAWDATA_DIR` is recursively scanned by the `Data Organiser` actor for input raw data. The directory `CALIB_DATA_DIR`, which is by default within the pipeline installation directory, is also scanned by the `Data Organiser` actor to find any static calibrations that may be missing in your `DataSet(s)`. If required, the user may edit the directories `BOOKKEEPING_DIR`, `LOGS_DIR`, `TMP_PRODUCTS_DIR`, and `END_PRODUCTS_DIR`, which correspond to the directories where book-keeping files, logs, temporary products and end products are stored, respectively (see the Reflex User Manual for further details; [Forchi \(2012\)](#)).

Under the “Global Parameters” area of the workflow canvas, the user may set the `FITS_VIEWER` parameter to the command used for running his/her favourite application for inspecting FITS files. Currently this is set by default to `fv`, but other applications, such as `ds9`, `skycat` and `gaia` for example, may be useful for inspecting image data. Note that in general it is recommended to specify the full path to the visualization application.

By default the `EraseDirs` parameter is set to `false`, which means that no directories are cleaned before executing the workflow, and the recipe actors will work in Lazy mode (see Section 6.2.4), reusing the previous pipeline recipe outputs where input files and parameters are the same as for the previous execution, which saves considerable processing time. Sometimes it is desirable to set the `EraseDirs` parameter to `true`, which forces the workflow to recursively delete the contents of the directories specified by `BOOKKEEPING_DIR`, `LOGS_DIR`, and `TMP_PRODUCTS_DIR`. This is useful for keeping disk space usage to a minimum and will force the workflow to fully rereduce the data each time the workflow is run.

The parameter `RecipeFailureMode` controls the behaviour in case that a recipe fails. If set to `Continue`, the workflow will trigger the next recipes as usual, but without the output of the failing recipe, which in most of the cases will lead to further fails of other recipes without the user actually being aware of it. This mode might be useful for unattended processing of large number of datasets. If set to `Ask`, a pop-up window will ask whether the workflow should stop or continue. This is the default. Additionally, the `Stop` mode will stop the workflow execution immediately.

The parameter `GlobalPlotInteractivity` controls whether the interactive windows will appear for those windows which are *enabled* by default. The possible values are `true`, `false`. Take into account that some windows are disabled in the default configuration and therefore are not affected by this parameter.



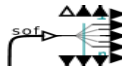


The parameter `ProductExplorerEnabled` controls whether the `ProductExplorer` actor will show its window or not. The possible values are `Enabled`, `Disabled` and `Triggered`. The later, recommended, means that the `ProductExplorer` actor will be shown only at the end of the workflow execution.

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6.2 Workflow Actors

6.2.1 Simple Actors

Simple actors have workflow symbols that consist of a single (rather than multiple) green-blue rectangle. They may also have a logo within the rectangle to aid in their identification. The following actors are simple actors:

- 
 - The Data Organiser actor.
- 
 - The Data Set Chooser actor.
- 
 - The Fits Router actor
- 
 - The Product Renamer actor.
- 
 - The Product Explorer actor.

Access to the parameters for a simple actor is achieved by right-clicking on the actor and selecting `Configure Actor`. This will open an “Edit parameters” window. Note that the `Product Renamer` actor is a jython script (Java implementation of the Python interpreter) meant to be customised by the user (by double-clicking on it).

6.2.2 Composite Actors

Composite Actors have workflow symbols that consist of multiple-layered green-blue rectangles. They generally do not have a logo within the rectangle. A Composite Actor represents a combination of more Simple or Composite Actors which hides over-complexity from the user in the top-level workflow. In the FORS2 workflow, the following actors are composite actors:

- 
 - The Initialise actor.
- 
 - The Initialise Current DataSet actor.
- 
 - The MasterBias actor.

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-  - The `ForsCalib` actor.
-  - The `ResponseCurve` actor.
-  - The `ForsScience` actor.
-  - The `Close DataSet` actor.

Composite Actors may also be expanded for inspection. To do this, right-click on the actor and select `Open Actor`, which will expand the Composite Actor components in a new `Reflex` canvas window. If the Composite Actor corresponds to a pipeline recipe, then the corresponding `RecipeExecutor` actor will be present as a Simple Actor, and its parameters are accessible as for any other Simple Actor. Alternatively you may still find Composite Actors, on which you need to repeat the first step to access the `Recipe Executor`.

6.2.3 Recipe Execution within Composite Actors

The FORS2 workflow contains Composite Actors to run pipeline recipes. This is in the most simple case due to the `SoF Splitter/SoF Accumulator`⁷, which allow to process calibration data from different settings within one given `DataSet` (e.g. lamp frames taken with different slits/masks). More complex Composite Actors contain several actors (e.g. `Recipe Executor`).

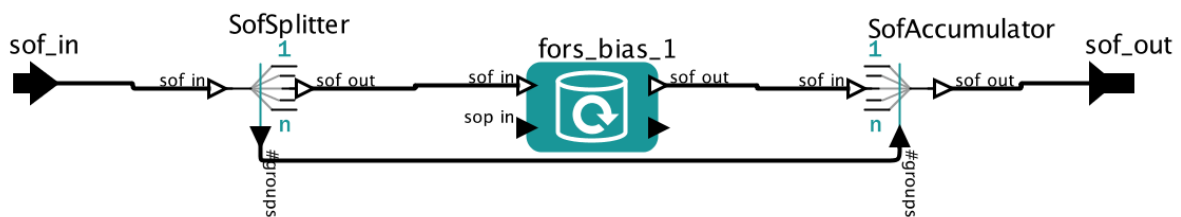


Figure 6.1: This is the window you get when you choose `Open Actor` for the Composite Actor `MasterBias`. This is the most simple case for a Composite Actor. Using `Configure Actor` on `fors_bias_1` gives you Fig. 6.2.

The central elements of any `Reflex` workflow are the `RecipeExecutor` actors that actually run the recipes. One basic way to embed a `RecipeExecutor` in a workflow is shown in Fig 6.1, which is the most simple version of a Composite Actor. The `RecipeExecutor` is preceded by an `SofSplitter`, and followed by an `SofAccumulator`. The function of the `SofSplitter` is to investigate the incoming `SoFs`, sort them by “purpose”, and create separate `SoFs` for each purpose. The `RecipeExecutor` then processes each of

⁷ `SoF` stands for `Set of Files`, which is an ASCII file containing the name (and path) of each input file and its category (e.g. `BIAS`).

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Table 6.1: The FORS2 pipeline actors and their contents

actor	recipes	description
MasterBias	fors_bias	create master bias
ForsCalib	fors_pmos_calib	create master flat, determine coefficients for wave-length calibration and correction of spatial distortion
Response Curve	fors_pmos_science	determine response function
ForsScience	fors_pmos_science	reduce science data

the SoFs independently (unless they are actually the same files). Finally, the `SofAccumulator` packs all the results into a single output SoF. The direct relation between the `SofSplitter` and `SofAccumulator` is used to communicate the number of different SoFs created by the `SofSplitter`. A workflow will only work as intended if the purpose of all the files a recipe needs as input is identical. The only exception to this rule is that a purpose can also be “default”. In this case, the file is included in any output SoF created by the `SoFsplitter` and `SofAccumulator`.

The reason for this scheme is best explained by an example. For a complex `DataSet`, the `Data Organiser` might have selected a large number of individual raw lamp frames (arc and flat field). The different lamp frames are to be used to calibrate different frames, e.g. the science frames and the standard star frames. The `Data Organiser` determines and records this “purpose” of each lamp frame, and this information is included in the `DataSet` and each SoF created from this `DataSet`. The `FitsRouter` directs all raw lamp frames to the `ForsCalib` Composite Actor. The `SofSplitter` then creates SoFs, one for the lamp frames to be used for the science frames, and (probably) separate ones for the lamp frames to be used for the standard star observations. The `fors_calib` recipe creates one master flat field (and other products) for each SoF, and the `SofAccumulator` then creates a SoF that contains all the products.

A `RecipeExecutor` actor is used in the workflow to run a single FORS2 pipeline recipe (e.g. in the `MasterBias` actor the recipe `fors_bias` is executed). In order to configure the `RecipeExecutors`, one has to first use `Open Actor` to get to the level of the recipe executors (see Fig. 6.1).

In Figure 6.2 we show the “Edit parameters” window for a typical `RecipeExecutor` actor, which can be displayed by right-clicking on the actor and selecting `Configure Actor`. In the following we describe in more detail the function of some of the parameters for a `RecipeExecutor` actor:

- The “recipe” parameter states the FORS2 pipeline recipe which will be executed.
- The “mode” parameter has a pull-down menu allowing the user to specify the execution mode of the actor. The available options are:
 - Run: The pipeline recipe will be executed, possibly in Lazy mode (see Section 6.2.4). This option is the default option.
 - Skip: The pipeline recipe is not executed, and the actor inputs are passed to the actor outputs.
 - Disabled: The pipeline recipe is not executed, and the actor inputs are not passed to the actor outputs.
- The “Lazy Mode” parameter has a tick-box (selected by default) which indicates whether the `RecipeExecutor` actor will run in Lazy mode or not. A full description of Lazy mode is provided in Sect. 6.2.4.

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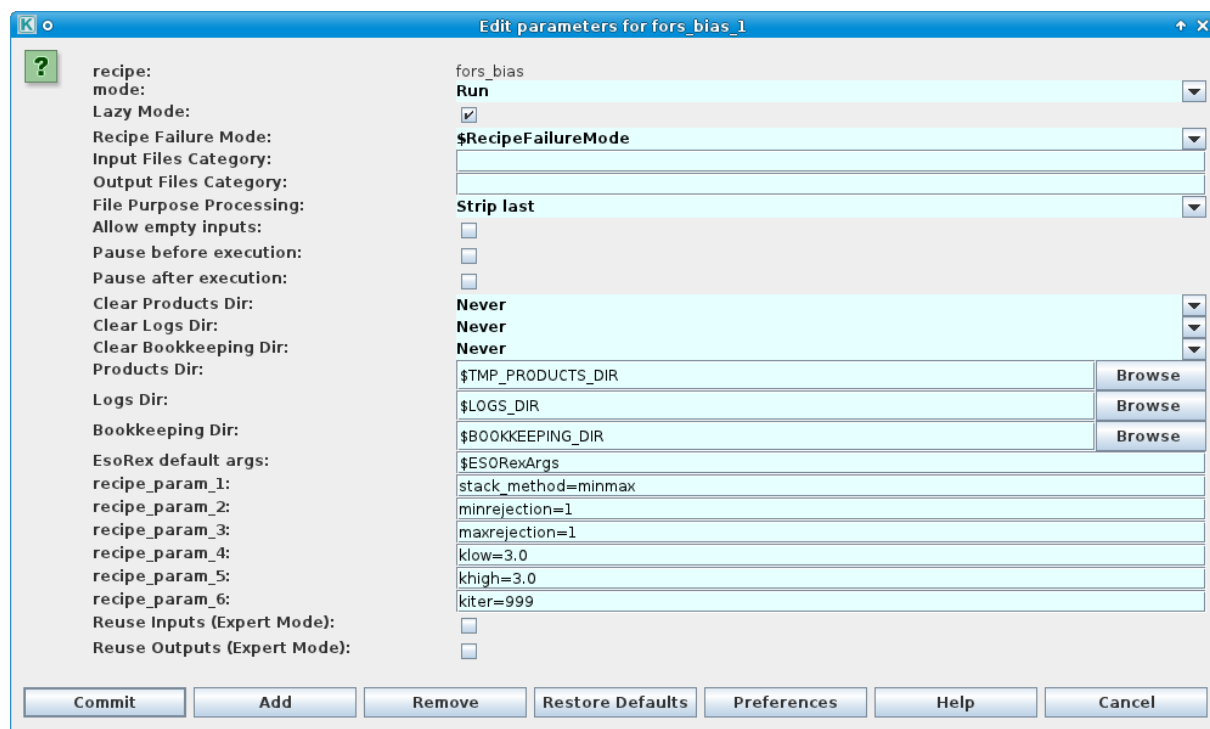


Figure 6.2: The “Edit parameters” window for a typical `RecipeExecutor` actor, the `fors_bias_1` actor which runs the `fors_bias` pipeline recipe.

- The “Recipe Failure Mode” parameter has a pull-down menu allowing the user to specify the behaviour of the actor if the pipeline recipe fails. The available options are:
 - Stop: The actor issues an error message and the workflow stops.
 - Continue: The actor creates an empty output and the workflow continues.
 - Ask: The actor displays a pop-up window and asks the user whether he/she wants to continue or stop the workflow. This option is the default option.
- The set of parameters which start with “recipe param” and end with a number or a string correspond to the parameters of the relevant FORS2 pipeline recipe. By default in the `RecipeExecutor` actor, the pipeline recipe parameters are set to their pipeline default values. If you need to change the default parameter value for any pipeline recipe, then this is where you should edit the value⁸. For more information on the FORS2 pipeline recipe parameters, the user should refer to the FORS2 pipeline user manual (Izzo et al. 2012⁹).

The description of the remainder of the `RecipeExecutor` actor parameters are outside the scope of this tutorial, and the interested user is referred to the Reflex User Manual for further details (Forchì 2012). Any

⁸Some of the pipeline parameters are read from the `GRISM_TABLES`, which contain grism-specific parameters. These cannot be changed here.

⁹Available at <ftp://ftp.eso.org/pub/dfs/pipelines/fors/fors-pipeline-manual-5.2.pdf>

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changes that you make in the “Edit parameters” window may be saved in the workflow by clicking the Commit button when you have finished.

6.2.4 Lazy Mode


By default, all recipe executor actors in a pipeline workflow are “Lazy Mode” enabled. This means that when the workflow attempts to execute such an actor, the actor will check whether the relevant pipeline recipe has already been executed with the same input files and with the same recipe parameters. If this is the case, then the actor will not execute the pipeline recipe, and instead it will simply broadcast the previously generated products to the output port. The purpose of the Lazy mode is therefore to minimise any reprocessing of data by avoiding data rereduction where it is not necessary.

One should note that the actor’s Lazy mode depends on the contents of the directory specified by `BOOKKEEPING_DIR` and the relevant FITS file checksums. Any modification to the directory contents and/or the file checksums will cause the corresponding actor when executed to run the pipeline recipe again, thereby rereducing the input data.

The forced rereduction of data at each execution may of course be desirable. To force a rereduction of all data for all `RecipeExecutor` actors in the workflow (i.e. to disable Lazy mode for the whole workflow), set the `EraseDirs` parameter under the “Global Parameters” area of the workflow canvas to `true`. This will then remove all previous results as well. To force a rereduction of data for any single `RecipeExecutor` actor in the workflow (which will be inside the relevant composite actor), right-click the `RecipeExecutor` actor, select `Configure Actor`, and uncheck the Lazy mode parameter tick-box in the “Edit parameters” window that is displayed. If the Lazy mode is switched off for an actor, all subsequent actors that use products from that one will also reprocess the data, as they see new products.

6.3 Workflow Steps

6.3.1 Step 1: Data Organisation And Selection

On clicking the  button on the `Reflex` canvas, the workflow will highlight and execute the `Initialise` actor, which among other things will clear any previous reductions if required by the user (see Section 6.1).

1. The `DataOrganiser` (DO) is the first crucial component of a `Reflex` workflow. The DO takes as input `RAWDATA_DIR` and `CALIB_DATA_DIR` and it detects, classifies, and organises the files in these directories and any subdirectories. The output of the DO is a list of “DataSets”. A `DataSet` is a special Set of Files (SoF). A `DataSet` contains one or several science (or calibration) files that should be processed together, and all files needed to process these data. This includes any calibration files, and in turn files that are needed to process these calibrations. Note that different `DataSets` might overlap, i.e. some files might be included in more than one `DataSet` (e.g. BIAS files).

A `DataSet` lists three different pieces of information for each of its files, namely 1) the file name (including the path), 2) the file category, and 3) a string that is called the “purpose” of the file. The DO uses

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OCA¹⁰ rules to find the files to include in a DataSet, as well as their categories and purposes. The file category identifies different types of files. A category could for example be `FLAT_PMO5`, `LAMP_PMO5` or `SCIENCE_PMO5`. The purpose of a file identifies the reason why a file is included in a DataSet. The syntax is `action_1/action_2/action_3/ ... /action_n`, where each `action_i` describes an intended processing step for this file. The actions are defined in the OCA rules and contain the recipe together with all file categories required to execute it (and predicted products in case of calibration data). For example, a workflow might include two actions `BIAS` and `CAL_PMO5`. The former creates a master bias from raw biases, and the later creates (among other products) a master flat from raw flats. The `CAL_PMO5` action needs raw lamp frames (arc and flat field) and the master bias as input. In this case, these biases will have the purpose `BIAS/CAL_PMO5`. The same DataSet might also include biases with a different purpose, e.g. `BIAS/SCIENCE_PMO5`. Irrespective of their purpose the file category for all these biases will be `BIAS`.

2. Next the `DataSet Chooser` displays the DataSets available in the “Select Data Sets” window, activating a vertical scroll bar on the right if necessary (see Figure 4.3). Sometimes you will want to reduce a subset of these DataSets rather than all DataSets, and for this you may individually select (or de-select) DataSets for processing using the tick boxes in the first column, and the buttons `Select All` and `Deselect All` at the bottom left.

You may also highlight a single DataSet in blue by clicking on the relevant line. If you subsequently click on `Inspect Highlighted`, then a “Select Frames” window will appear that lists the set of files that make up the highlighted DataSet including the full filename and path for each file, the file category (from the FITS header), and a selection tick box in the right column (see Figure 6.3). The tick boxes allow you to edit the set of files in the DataSet which is useful if it is known that a certain calibration frame is of poor quality (e.g: a poor raw flat-field frame). The list of files in the DataSet may also be saved to disk as an ASCII file by clicking on `Save As` and using the file browser that appears.

By clicking on the line corresponding to a particular file in the “Select Frames” window, the file will be highlighted in blue, and the file FITS header will be displayed in the text box on the right (see Figure 6.3), allowing a quick inspection of useful header keywords. If you then click on `Inspect`, the workflow will open the file in the selected FITS viewer application defined by the workflow parameter `FITS_VIEWER`.

To exit from the “Select Frames” window, click `Continue`, and to exit from the “Select DataSets” window, click either `Continue` in order to continue with the workflow reduction, or `Stop` in order to stop the workflow.

The categories and purposes of raw files are set by the DO, whereas the categories and purpose of products generated by recipes are set by the `RecipeExecuter` (see Sect. 6.2.3). The file categories are used by the `FitsRouter` to send files to particular processing steps or branches of the workflow (see below). The purpose is used by the `SofSplitter` and `SofAccumulator` to generate input SoFs for the `RecipeExecuter`. Note that while the DO includes files into a DataSet for a reason, and records this reason as the “purpose” of the file, the workflow itself can use these files in a different manner. The `SofSplitter` and `SofAccumulator` accept several SoFs as simultaneous input. The `SofAccumulator` creates a single output SoF from the inputs, whereas the `SofSplitter` creates a separate output SoF for each purpose.

¹⁰OCA stands for OrganisationClassificationAssociation and refers to rules, which allow to classify the raw data according to the contents of the header keywords, organise them in appropriate groups for processing, and associate the required calibration data for processing. They can be found in the directory `<install_dir>/share/esopipes/<pipeline-version>/reflex/`, carrying the extension `.oca`

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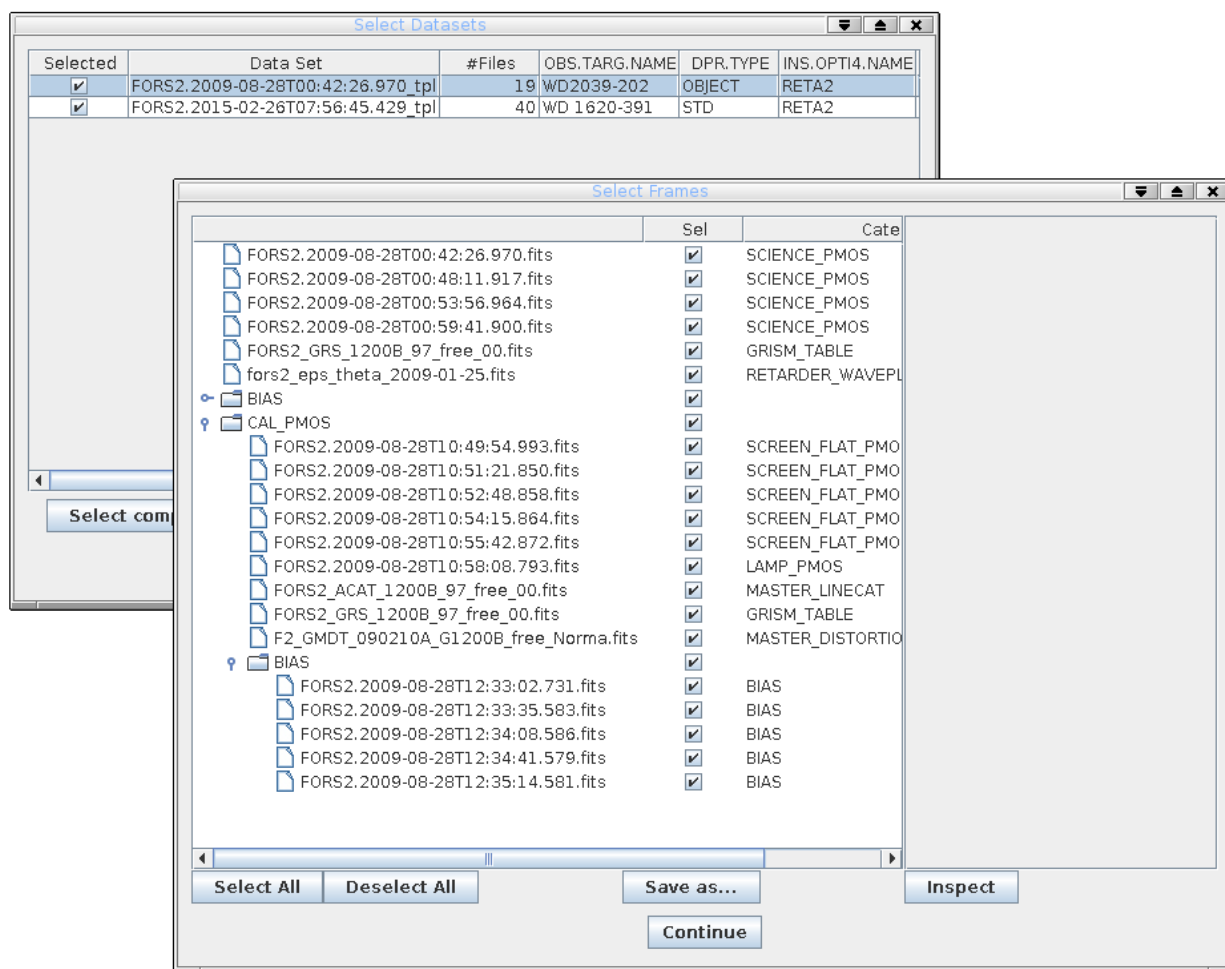


Figure 6.3: The “Select Frames” window with a single file from the current Data Set highlighted in blue, and the corresponding FITS header displayed in the text box on the right. Hidden partially behind the “Select Frames” window is the “Select DataSets” window with the currently selected DataSet highlighted in blue.

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6.3.2 Step 2: Creation Of Master Calibration Files

In this step of the workflow, the following FORS2 recipes are executed in the order listed below. Please refer to the FORS2 pipeline user manual (Izzo et al. 2012: Sections 9 and 10) for the details of each recipe and the algorithms employed:

1. The `MasterBias` actor will execute the FORS2 pipeline recipe `fors_bias` in order to create a combined master bias frame from the set of raw bias frames
2. The `ForsPmosCalib` actor will execute the FORS2 pipeline recipe `fors_pmos_calib` in order to create from the set of raw flat and arc frames a combined master flat frame as well as coefficients for wavelength calibration and correction of spatial distortions.

6.3.3 Step 3: Science Reduction

The `ForsPmosScience` actor will execute the FORS2 pipeline recipe `fors_pmos_science` to apply sky subtraction and extract the spectra. Please refer to the FORS2 pipeline user manual (Izzo et al. 2012: Sections 9 and 10) for the details of this recipe and the extraction algorithms employed.

6.3.4 Step 4: Output Organisation

After having processed the input data for a `DataSet`, the workflow highlights and executes the `Product Renamer` actor, which, by default, will copy the defined final products of the `ForsScience` actor to the directory specified by `END_PRODUCTS_DIR` and rename them with names derived from the values of certain FITS header keywords. Specifically, final products are renamed by default with names of the form `<HIERARCH.ESO.OBS.NAME>_<HIERARCH.ESO.PRO.CATG>.fits`, with `<HIERARCH.ESO.OBS.NAME>` and `<HIERARCH.ESO.PRO.CATG>` representing the values of the corresponding FITS header keywords (`<HIERARCH.ESO.OBS.NAME>` is the name of the OB and `<HIERARCH.ESO.PRO.CATG>` is the category of the product file). These names are fully configurable by right-clicking on the `Product Renamer` actor, selecting `Configure Actor`, and then editing the string as appropriate. In some cases the keyword `<HIERARCH.ESO.OBS.TARG.NAME>` (target name) may be more useful than `<HIERARCH.ESO.OBS.NAME>`.

For PMOS data the final products that are copied and renamed are (for better readability we replace `<HIERARCH.ESO.OBS.NAME>` by `<OB_NAME>`):

- **1-dimensional extracted spectra** (`<OB_NAME>_REDUCED_*`, created only if spectra are identified and can be extracted).

The individual spectra are provided as rows in a FITS file with as many extensions as input scientific exposures. The correspondence between these rows and the 2-dimensional frames and/or slit identifications can be obtained from `<OB_NAME>_OBJECT_TABLE_SCI_PMOS.fits`. All extracted spectra have the same format.

- `<OB_NAME>_REDUCED_SCI_PMOS.fits` spectra
- `<OB_NAME>_REDUCED_ERROR_SCI_PMOS.fits` 1σ error of spectra

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- `<OB_NAME>_REDUCED_SKY_SCI_PMOS.fits` fitted sky spectra
- **1-dimensional polarization spectra** (same format as 1-dimensional extracted spectra)
 - X may be any of `ANGLE`, `I`, `L`, `Q`, `U`, `V` according to the convention described in [Izzo et al. \(2012\)](#)
 - `<OB_NAME>_REDUCED_X_SCI_PMOS.fits` extracted polarisation signals from the object-sources.
 - `<OB_NAME>_REDUCED_ERROR_X_SCI_PMOS.fits` 1σ errors of the extracted polarisation signals
- `<OB_NAME>_OBJECT_TABLE_POL_SCI_PMOS.fits` table with position information for detected spectra, with the information from the extensions included within one table.
- **2-dimensional wavelength calibrated and distortion corrected frames** (`<OB_NAME>_MAPPED_*`, with as many extensions as input scientific exposures)
 - `<OB_NAME>_MAPPED_ALL_SCI_PMOS.fits` frame without sky subtraction
 - `<OB_NAME>_MAPPED_SCI_PMOS.fits` frame, sky-subtracted
 - `<OB_NAME>_MAPPED_SKY_SCI_PMOS.fits` frame with fitted sky background

If **sky alignment** is requested (`skyalign`≥0) the following products are provided in addition to the ones listed above:

- `<OB_NAME>_DISP_COEFF_SCI_PMOS.fits` adjustment of the input `DISP_COEFF_PMOS` table
- `<OB_NAME>_SKY_SHIFTS_SLIT_SCI_PMOS.fits` table with sky line shifts
- `<OB_NAME>_WAVELENGTH_MAP_SCI_PMOS.fits` wavelength map adjusted for sky line shifts

The following actors in this step of the workflow are concerned with the termination of the data flow for the current DataSet and will highlight briefly as they are executed.

Finally, the `Product Explorer` window will appear as shown in Fig. 4.4 with a list of datasets on the left menu. By unfolding the menu under each dataset, all the renamed products appear, and if one is interested in the files, including all intermediate steps, that are used to produce that final product, just click on it and a dependency tree will show the whole reduction chain.

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7 Frequently Asked Questions

- **The error window fills the whole screen - how can I get to the `Continue`/`Stop` buttons?**

Press the `Alt` key together with your left mouse button to move the window upwards and to the left. At the bottom the `Continue`/`Stop` buttons will be visible. This bug is known but a fix has not yet been found.

- **Where are my intermediate pipeline products?** Intermediate pipeline products are stored in the directory `<TMP_PRODUCTS_DIR>` (defined on the workflow canvas, under Setup Directories) and organised further in directories by pipeline recipe.
- **Can I use different sets of bias frames to calibrate my flat frames and science data?** Yes. In fact this is what is currently implemented in the workflow(s). Each file in a DataSet has a purpose attached to it (Forchì (2012)). It is this purpose that is used by the workflow to send the correct set of bias frames to the recipes for flat frame combination and science frame reduction, which may or may not be the same set of bias frames in each case.
- **Can I run Reflex from the command line?** Yes, use the command:

```
esoreflex -n <workflow_path>/<workflow>.xml
```

The `-n` option will set all the different options to Kepler and the workflows that avoid any graphical display (including pipeline interactive windows).

It is possible to specify workflow variables (those that appear in the workflow canvas) in the command line. For instance, to set the raw data directory can be done with this command:

```
esoreflex -n -RAW_DATA_DIR <raw_data_path> <workflow>.xml
```

You can see all the command line options with the command `./esoreflex -h`.

Note that this mode is not fully supported, and the user should be aware that the path to the workflow must be absolute and even if no GUI elements are shown, it still requires a connection to the window manager.

- **How can I add new actors to an existing workflow?** You can drag and drop the actors in the menu on the left of the Reflex canvas. Under `Eso-reflex -> Workflow` you may find all the actors relevant for pipeline workflows, with the exception of the recipe executor. This actor must be manually instantiated using `Tools -> Instantiate Component`. Fill in the “Class name” field with `org.eso.RecipeExecutor` and in the pop-up window choose the required recipe from the pull-down menu. To connect the ports of the actor, click on the source port, holding down the left mouse button, and release the mouse button over the destination port. Please consult the Reflex User Manual (Forchì (2012)) for more information.
- **How can I broadcast a result to different subsequent actors?** If the output port is a multi-port (filled in white), then you may have several relations from the port. However, if the port is a single port (filled in black), then you may use the black diamond from the toolbar. Make a relation from the output port to the

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diamond. Then make relations from the input ports to the diamond. Please note that you cannot click to start a relation from the diamond itself. Please consult the Reflex User Manual ([Forchi \(2012\)](#)) for more information.

- **How can I run manually the recipes executed by Reflex?** If a user wants to re-run a recipe on the command line he/she has to go to the appropriate `reflex_book_keeping` directory, which is generally `reflex_book_keeping/<workflow>/<recipe_name>_<number>`. There, subdirectories exist with the time stamp of the recipe execution (e.g. 2013-01-25T12:33:53.926/). If the user wants to re-execute the most recent processing he/she should go to the `latest` directory and then execute the script `cmdline.sh`. Alternatively, to ensure that the path to `esorex` is the correct one, the user can execute

```
ESOREX_CONFIG="INSTALL_DIR/etc/esorex.rc"
INSTALL_DIR/bin/esorex --recipe-config=<recipe>.rc <recipe> data.sof
```

where `INSTALL_DIR` is the directory where Reflex and the pipelines were installed.

If a user wants to re-execute in the command line a recipe that used a given raw frame, the way to find the proper `data.sof` in the bookkeeping directory is via `grep <raw_file> */data.sof`. Afterwards the procedure is the same as before.

If a recipe is re-executed with the command explained before, the products will appear in the directory from which the recipe is called, and not in the `reflex_tmp_products` or `reflex_end_products` directory, and they will not be renamed. This doesn't happen if you use the `cmdline.sh` script.

- **If I enter "-" into an empty integer parameter it is automatically completed to "-1". Why?**

The parameters are validated for correctness according to their type (e.g. string, integer, float). In the case of an integer or float parameter "-" alone is considered an invalid input and is therefore automatically completed to "-1". This is part of the validation of input done by the WxPython library.

- **Can I reuse the bookkeeping directory created by previous versions of the pipeline?**

In general no. In principle, it could be reused if no major changes are in the pipeline. However there are situations in which a previously created bookkeeping directory will cause problems due to pipeline versions incompatibility. This is specially true if the parameters of the pipeline recipes have changed. In that case, please remove completely the bookkeeping directory.

7.1 FORS specific questions

- **There are no extracted spectra - why?**

While this may happen for observations of extremely faint targets the most common case for this behaviour are PMOS observations from the lower chip, as the main target is usually placed close to the centre of the field-of-view, which is on the upper chip.

- **Does the pipeline combine different detectors chips into a common product?**

No. The spectroscopic pipeline and Reflex workflow works only on files from the same detector chip. Files from different detectors must not be mixed in the same `sof`.

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8 Troubleshooting



Figure 8.1: *TheDataOrganizer* interactive window reports an error “:No DataSets have been created, check the data set and the OCA rules.”.

1. I downloaded the data from the ESO archive, put them into a new directory, tried to run Reflex on them, but

- (a) it crashes

The current release of FORS includes some additional data in the static calibration frames. The recipes would chock if this data is not present. However, the ESO archive with CalSelector will associate calibration data which is old and Reflex will pick the files either from the installed pipeline static data or from the CalSelector in a non-deterministic way. In order to solve the issue, remove the static calibration data downloaded from the archive (all the files starting with M.FORS2).

This may happen if one of the files was downloaded only partially (check for a file with the extension `fits.Z.part`. You will have to download that file again in order to have an uncorrupted file (and remove the partial one).

- (b) The DataOrganiser fails with the error message “:No DataSets have been created, check the data set and the OCA rules.”(see Figure 8.1.)

This error may be due to the fact that the data provided by the ESO archive are compressed (`<filename>.fits.Z`). Please remember to uncompress the data before running the workflow in Reflex.

Also, please remember that the FORS2 workflow supports only spectro-polarimetric data (PMOS). It is possible that your data consists entirely of IMG/IPOL/LSS/MOS/MXU observations, in which case the Data Organiser actor will not construct any DataSets, showing the mentioned error message.

2. The “Select DataSets” window displays my DataSets, but some/all of them are greyed out. What is going on?

If a DataSet in the “Select DataSets” window is greyed out, then it means that the DataSet which was constructed is missing some key calibration(s) (i.e. the DataSet is incomplete). To find out what calibration(s) are missing from a greyed out DataSet, click on the DataSet in question to highlight it in blue, and then click on the button `Inspect Highlighted`. The “Select Frames” window that appears will

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report the category of the calibration products that are missing (e.g. MASTER_BIAS). From this the user has then to determine the missing raw data (in this case bias frames). If static calibrations are missing the mechanism unfortunately does not work, but such data should be found by `reflex` in `<install_directory>/calib/<pipeline_version>/cal`

3. The pipeline fails with the error message “The wavelength solution at row <number> does not increase monotonically, which is physically impossible. Try with new parameters”.

Non-monotonic dispersion relations are often due to spurious detections. In such cases try to decrease `wdegree` and/or increase `peakdetection`. A further possibility is to decrease `wradius`.

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